

RESEARCH ARTICLE

Open Access

Mangotoxin production of *Pseudomonas syringae* pv. *syringae* is regulated by MgoA

Víctor J Carrión^{1,3,4}, Menno van der Voort³, Eva Arrebola², José A Gutiérrez-Barranquero^{1,5}, Antonio de Vicente¹, Jos M Raaijmakers^{3,4} and Francisco M Cazorla^{1*}

Abstract

Background: The antimetabolite mangotoxin is a key factor in virulence of *Pseudomonas syringae* pv. *syringae* strains which cause apical necrosis of mango trees. Previous studies showed that mangotoxin biosynthesis is governed by the *mbo* operon. Random mutagenesis led to the identification of two other gene clusters that affect mangotoxin biosynthesis. These are the *gacS/gacA* genes and *mgo* operon which harbors the four genes *mgoBCAD*.

Results: The current study shows that disruption of the nonribosomal peptide synthetase (NRPS) gene *mgoA* resulted in loss of mangotoxin production and reduced virulence on tomato leaves. Transcriptional analyses by qPCR and promoter reporter fusions revealed that *mbo* expression is regulated by both *gacS/gacA* and *mgo* genes. Also, expression of the *mgo* operon was shown to be regulated by *gacS/gacA*. Heterologous expression under the native promoter of the *mbo* operon resulted in mangotoxin production in non-producing *P. syringae* strains, but not in other *Pseudomonas* species. Also introduction of the *mbo* and *mgo* operons in nonproducing *P. protegens* Pf-5 did not confer mangotoxin production but did enhance transcription of the *mbo* promoter.

Conclusions: From the data obtained in this study, we conclude that both *mbo* and *mgo* operons are under the control of the *gacS/gacA* two-component system and that the MgoA product acts as a positive regulator of mangotoxin biosynthesis.

Keywords: Antimetabolite toxin, *mgo* operon, GacS/GacA, Plant-microbe interaction

Background

Pseudomonas syringae is one of the most ubiquitous plant pathogens, causing various economically important diseases [1]. The present study focuses on *P. syringae* pv. *syringae* UMAF0158 (CECT 7752) which causes apical necrosis of mango [2,3]. The antimetabolite mangotoxin is a key virulence factor of strain UMAF0158 [4,5]. This toxin is produced in the early exponential growth phase and inhibits ornithine N-acetyl transferase, a key enzyme belonging to the ornithine/arginine biosynthetic pathway [2].

Random mini-Tn5 mutagenesis followed by cloning, sequencing and heterologous expression recently led to the identification of the gene cluster that governs mangotoxin biosynthesis [6]. The *mbo* operon (**mangotoxin biosynthetic**

operon) is composed of six genes, *mboABCDEF*. Disruption of each of these genes resulted in mangotoxin deficient mutants and constitutive expression of the *mbo* operon in non-mangotoxin producing *P. syringae* strains conferred mangotoxin production [6]. Screening of the random mutant library also led to the identification of several other genes that may be involved in the regulation of mangotoxin biosynthesis [4]. These included the *gacS/gacA* genes and the so-called **mangotoxin generating operon** *mgo* [6,7].

The GacS/GacA two-component regulatory system is highly conserved in Gram-negative bacteria and is involved in a variety of functions, including pathogenicity [8], quorum sensing [9,10], secondary metabolite production [11-14] and biofilm formation [15-17]. In *Pseudomonas syringae*, the GacS/GacA two-component system regulates the production of the phytotoxins syringomycin and syringopeptin [18-20], tabtoxin [21,22] and phaseolotoxin [23]. In *P. syringae* pv. *tomato* DC3000, GacS/GacA regulate the *hrpR*, *hrpS*, and *hrpL* genes, which are required for the activation of the Hrp

* Correspondence: cazorla@uma.es

¹Departamento de Microbiología, Facultad de Ciencias, Instituto de Hortofruticultura Subtropical y Mediterránea "La Mayora"-Universidad de Málaga-Consejo Superior de Investigaciones Científicas (IHSM-UMA-CSIC), Universidad de Málaga, Campus de Teatinos, 29071 Málaga, Spain
Full list of author information is available at the end of the article

type III secretion and effector genes [24,25]. However, in *P. syringae* pv. *syringae* B728a, GacA appears not to be required for *hrp* gene expression [25].

The *mgo* operon is composed of four genes, *mgoBCAD* [4,7]. Mutants in each gene belonging to the *mgo* operon showed an alteration (*mgoB* mutant) or lack of mangotoxin production (*mgoC*, *mgoA* and *mgoD* mutants). These genes encode for different hypothetical proteins with predicted domains for a haem oxygenase (MgoB), a p-aminobenzoate N-oxygenase (MgoC), a nonribosomal peptide synthetase (MgoA), and a polyketide cyclase/dehydrase or lipid transporter (MgoD) [4,7]. The predicted amino acid sequence of MgoA suggests only one amino acid activation module and 14 conserved domains, including aminoacyl adenylation, condensation, thiolation, and additional reduction domains [4]. Genes homologous to the *mgo* operon have been found in the genomes of most *Pseudomonas* spp., with the exception of *P. protegens* Pf-5 and CHAO [26,27]. Recent studies on the *pvf* gene cluster in *P. entomophila*, a homologue of the *mgo* operon, suggested that it affects virulence [28]. Almost all the fluorescent *Pseudomonas* spp. lack the *mbo* operon [29,30], but the *mgo* operon is conserved in all of them (except *P. protegens* Pf-5) [4,7,26-28]. To date, however, the functions of *mgo* operon are yet unknown.

The overall objective of this study was to get insight into the role of the *mgo* operon in regulation of mangotoxin production in *P. syringae* pv. *syringae* UMAF0158 and unravel the interplay between *mgo*, *mbo* and the *gacS/gacA* two-component regulatory system.

Methods

Bacterial strains and culture conditions

The wild type strain *P. syringae* pv. *syringae* UMAF0158 (CECT 7752) and the collection of selected derivative mutants used in this study (Table 1) were grown on *Pseudomonas* agar F (Difco) plates, in liquid King's medium B (KMB) [31] or in *Pseudomonas* minimal medium (PMS) [32] at 28°C. *Escherichia coli* strain DH5α was used as a host for plasmid complementation experiments. It was routinely grown on Luria-Bertani (LB) plates or in LB broth at 37°C. Antibiotics for selection of *P. syringae* pv. *syringae* UMAF0158 and *E. coli* derivatives were ampicillin (100 mg L⁻¹), kanamycin (50 mg L⁻¹), gentamycin (30 mg L⁻¹) or tetracycline (25 mg L⁻¹).

Mangotoxin production assay

Antimetabolite toxin production was assayed by the indicator technique previously described [32]. Briefly, a double layer of the indicator microorganism *E. coli* CECT 831 was prepared; after solidification, the *P. syringae* pv. *syringae* strains to be tested were stab-inoculated. The plates were

initially incubated at 22°C for 24 h, and then at 37°C for an additional 24 h [2]. To evaluate mangotoxin activity, the same plate bioassay was carried out with the addition of 100 μl of a 6 mM solution of *N*-acetyl-ornithine or L-ornithine to the double layer of *E. coli* [2]. To determine growth characteristics of representative strains, the wild type mangotoxin-producing *P. syringae* pv. *syringae* UMAF0158 and derivatives mutants in *mboA*, *mgoA* and *gacA* genes were used to obtain initial cultures in 10 ml of LB broth. The bacterial strains were grown during 24 h at 28°C to prepare an optimal bacterial inoculum with an optical density of 0.8 at 600 nm (approximately 10⁹ cfu ml⁻¹). One ml from these bacterial inocula was used to inoculate 100 ml of PMS broth. The cultures were incubated at 22°C under orbital shaking at 150 rpm until the stationary phase. Samples were collected every 6 or 12 h to monitor the bacterial growth. Bacterial cfu per sample were determined by 10-fold serial dilutions on KMB plates. At the same time, the mangotoxin production assessment was performed by a cell-free filtrate dilution sequence at 50%. The mangotoxin production is measured using arbitrary units, which can be defined as the relative toxic volume of cell free filtrates of liquid cultures, which produces an inhibition halo of 18 mm in diameter under standard assay conditions [2]. The methodology presented a detection threshold of 0.5 toxic units, due to the diameter of the wells where the cell-free filtrate were deposited (9 mm).

Complementation experiments

DNA fragments of approximately 7 kb containing the *mgo* and *mbo* operons, including the promoter and terminator regions, were obtained by PCR using specific primers (Additional file 1: Table S1) and high fidelity polymerase (Phusion DNA polymerase, Finnzymes). The PCR amplification products were cloned in pGEM-T Easy (Promega), and the plasmids obtained were digested with *Xba*I for the *mgo* operon and with *Eco*RI and *Pst*I for the *mbo* operon. After the digestion, both operons fragment were obtained from gel with the NucleoSpin kit (GE Healthcare) and cloned into the correspondent shuttle vectors, pBBR1MCS-5 [36] for the *mgo* operon and pMP220 [37] for the *mbo* operon, which were digested, dephosphorylated (shrimp alkaline phosphatase; Promega), and purified with the NucleoSpin kit according to the manufacturer's instructions. *E. coli* DH5α was transformed with the plasmids obtained, by heat shock transformation [38], and transformed colonies were selected on LB agar plates supplemented with gentamicin (30 mg L⁻¹) in the case of pBBR1MCS-5 and tetracycline (25 mg L⁻¹) for pMP220. Plasmids with the *mgo* and *mbo* operon cloned were obtained (Table 1). Correct integration and orientation of the fragments was verified by PCR and restriction analysis of isolated plasmids (data not shown). The pLac-*mgoBCAD* construct was subsequently electroporated into the *mboA*, *mgoA* and *gacA*

Table 1 Bacterial strains and plasmids used in this study

Strain or plasmid	Relevant characteristics	Reference/source
Strains		
<i>E. coli</i>		
DH5α	<i>E. coli</i> [F' Φ 80lacZ Δ M15 Δ (lacZYA-argF)U169 deoR recA endA1 hsdR17 (rK-mK+)phoA supE44 lambda-thi-1]	[33]
CECT831	Indicator strain for mangotoxin production	CECT ^a
<i>P. syringae</i> pv. <i>syringae</i>		
B728a	Complete genome, non-mangotoxin producer	[34]
UMAF0158	Wild type, isolated from mango, mangotoxin producer, Nf ^r	[2]
<i>mboA</i> ^r	Derivative mutant of UMAF0158 by insertion in <i>mboA</i> , Km ^r , Nf ^r (named <i>mboA</i>)	[6]
Δ <i>mgoA</i>	Derivative mutant of UMAF0158 by deletion of <i>mgoA</i> , Nf ^r (named Δ <i>mgoA</i>)	[7]
2 β B7	miniTn5 mutant of UMAF0158 in <i>gacA</i> defective in mangotoxin, Km ^r , Nf ^r (named <i>gacA</i>)	[4]
3 α E10	miniTn5 mutant of UMAF0158 in <i>gacS</i> defective in mangotoxin, Km ^r , Nf ^r (named <i>gacS</i>)	[2]
3 γ H1	miniTn5 mutant of UMAF0158, defective in mangotoxin production, Km ^r , Nf ^r	[2]
4 β A2	miniTn5 mutant of UMAF0158, defective in mangotoxin production, Km ^r , Nf ^r	[2]
5 α C5	miniTn5 mutant of UMAF0158, defective in mangotoxin production, Km ^r , Nf ^r	[2]
6 γ F6	miniTn5 mutant of UMAF0158, defective in mangotoxin production, Km ^r , Nf ^r	[2]
<i>P. protegens</i> Pf-5	Non mangotoxin producer, <i>mbo</i> and <i>mgo</i> operon absent	[35]
Plasmids		
pBBR1MCS-5	4.7 kb broad-host-range cloning vector, Gm ^r	[36]
pGEM-T	3.0 kb cloning vector, Ap ^r	Invitrogen
pGEM-TBCAD	<i>mgoBCAD</i> cloned in pGEM-T, Ap ^r	This study
pLac- <i>mgoBCAD</i>	<i>mgoBCAD</i> cloned in pBBR1MCS-5 downstream the <i>lacZ</i> promoter in the vector, <i>mgo</i> operon expression under its own and P _{LAC} promoter, Gm ^r	This study
pLac- <i>mboABCDEF</i>	<i>mboABCDEF</i> cloned in pBBR1MCS-5 downstream the <i>lacZ</i> promoter in the vector, <i>mbo</i> operon expression under its own and P _{LAC} promoter, Gm ^r	[6]
pLac- <i>mboFEDCBA</i>	<i>mboABCDEF</i> cloned in pBBR1MCS-5 in the opposite direction than the <i>lacZ</i> promoter in the vector, <i>mbo</i> operon expression under its own promoter, Gm ^r	[6]
pMP220	Promoter-probe vector containing a promoterless LacZ gene, Tet ^r	[37]
pMP- <i>mboABCDEF</i>	<i>mboABCDEF</i> cloned in promoter-probe vector containing a promoterless LacZ gene, <i>mbo</i> operon expression under its own promoter, Tet ^r	This study
pMP::P _{mbo}	pMP220 vector containing the <i>mbo</i> operon promoter, Tet ^r	[6]

^aCECT: Spanish Type Culture Collection, Spain.

mutants, and the wild-type strains *P. syringae* pv. *syringae* UMAF0158 and *P. protegens* Pf-5. The pMP-*mboABCDEF* construct was transformed in *P. protegens* Pf-5 which previously contain the pLac-*mgoBCAD*, therefore this bacteria finally harbored both operons, the *mgo* and *mbo* operon. Transformed cells were selected on KMB agar supplemented with correspondent antibiotics. The presence of the different plasmids was confirmed by PCR analysis with specific primers for pBBR1MCS-5 and pMP220 and plasmid profiling.

Virulence evaluation

The virulence of different mangotoxin producing or non-producing *P. syringae* pv. *syringae* strains were analyzed in detached tomato leaflets (*Solanum lycopersicum*

Mill.) cv. Hellfrucht Frühstamm maintained *in vitro* using Murashige and Skoog medium (MS, Sigma-Aldrich) [4,5]. Bacterial suspensions from exponentially growing cultures were adjusted to 10⁸ cfu ml⁻¹. The leaflets were inoculated by placing six 10 µl drops of the bacterial suspension on six different points on the same leaflet. Inoculations were then carried out by piercing through the droplets with a sterile entomological pin. The leaflets were maintained in MS media at 22°C and a 16:8-h light: dark photoperiod. Six tomato leaflets were used to evaluate each strain. Detached leaflets only inoculated with sterile distilled water were included in all experiments as a control. These experiments were repeated three times. The development of necrotic symptoms at the inoculation points (n = 108) was

determined after 10-day. The severity symptoms were evaluated by the analysis of the total necrotic area per leaflet induced by the inoculated strains after 10 days of incubation. For severity measurement, the necrotic areas of the inoculation points were digitally analyzed on the six leaflets, using the computer image software VISILOG 5.0 (Noesis Vision Inc.). At the same time, two inoculated leaflets were used to estimate the daily development of the total bacterial population. For that purpose, whole tomato leaflets were homogenized in sterile water and bacterial counts were determined plating by 10-fold serial dilutions on KMB plates. Bacterial growth inside the plant tissue was recorded after H₂O₂ leaf surface disinfection. Colony counts growth based on the typical morphology of *P. syringae* pv. syringae UMAF0158 were recorded after incubation at 28°C for 48 h.

Transcriptional analysis

From PMS cultures described above, cells from 2 ml cultures were collected and spun down at 12,000 rpm (1 min) from the wild type strain and the derivative mutants in *gacA* and *mgoA*. The cells were frozen in liquid N₂ and stored at -80°C. For the RNA isolations and cDNA synthesis, three biological replicates were used for each time point. For the transcriptional analyses, RNA was isolated from the frozen bacterial cells with Trizol reagent (Invitrogen), followed by DNase I (GE Healthcare) treatment. One µg of RNA was used for cDNA synthesis with Superscript III (Invitrogen) according to the manufacturer's protocol. For the real-time quantitative PCR (Q-PCR), conducted with the 7300SDS system from Applied Biosystems, the SYBR Green Core kit (Eurogentec) with a final concentration of 3.5 mM MgCl₂ was used according to the manufacturer's protocol. The concentration of the primers was optimized (400 nM final concentration for all of them), and a dissociation curve was performed to check the specificity of the primers. The primers used for the Q-PCR are listed in Additional file 1: Table S1. To correct for small differences in template concentration, *rpoD* was used as the reference housekeeping gene. The cycle in which the SYBR green fluorescence crossed a manually set cycle threshold (*C_T*) was used to determine transcript levels. For each gene, the threshold was fixed based on the exponential segment of the PCR curve. The *C_T* value of *mgoA* was corrected for the housekeeping gene *rpoD* as follows: $\Delta C_T = C_T(mgoA) - C_T(rpoD)$; the same formula was used for the other genes studied. The relative quantification (RQ) values were calculated by the following formula: RQ = $2^{-|\Delta C_T(\text{mutant}) - \Delta C_T(\text{wild type})|}$ [39,40]. Q-PCR analysis was performed in duplicate (technical replicates) on three independent RNA isolations (biological replicates).

β-galactosidase assays

To study the *mbo* operon expression in different genetic backgrounds, the *mbo* operon promoter (P_{*mbo*}) cloned into pMP220 [19] as previously described [6] was used. The derivative mutants in *mgoA*, *gacA* and *gacS* genes were transformed with plasmid pMP::P_{*mbo*} which contains the P_{*mbo*}. The plasmid pLac-*mgoBCAD* (harboring the *mgo* operon) was also used to complement the *mgoA*, *gacA* and *gacS* mutants and finally the β-galactosidase activity of P_{*mbo*} was measured. In order to evaluate the effect of the *mgo* operon on the activity of P_{*mbo*} *P. protegens* Pf-5 was used due to the absence of the two operons in its genome. First, *P. protegens* Pf-5 was transformed with the pMP::P_{*mbo*} and the promoter activity was measured, and secondly to measure the effect on the *mbo* operon transcription, this strain containing the plasmid pMP::P_{*mbo*} was also transformed with the plasmid pLac-*mgoBCAD* (*mgo* operon under pLac regulation). As a negative control the β-galactosidase activity was measured for the wild type strain *P. syringae* pv. syringae UMAF0158 and each strain used in this assay, transformed with empty vector pMP220. β-galactosidase activities were quantified by the Miller method [41]. Briefly, an overnight culture obtained as previously described in growth curve and toxins assay section were prepared. The samples were collected at 18 h, and the cells were harvested and suspended in assay buffer to eliminate any error in the detection of β-galactosidase activity due to the effects of different carbon sources present in the growth medium. The results presented are from three separate experiments, each conducted in triplicate.

Phylogeny of the *mgoA* gene

In order to identify the presence of the *mgoA* gene in the different genomes of *Pseudomonas* strains, the *mgoA* gene from *P. syringae* pv. syringae UMAF0158 was used in BLASTP [42] comparisons with whole genome sequences of *Pseudomonas* spp. available in the databases. Once the amino acid sequences of all the orthologous *mgoA* genes were obtained, the putative adenylation domains were identified using the PKS/NRPS Analysis Web-site (<http://nrps.igs.umaryland.edu/nrps>) [43]. Other adenylation domains of which the activated amino acid is already known were obtained from the database and from De Bruijn et al. [44]. Two phylogenetic analyses were done, the first was using the adenylation domain of all the NRPSs (328 residues) and the second was using the almost entire sequence of MgoA (1015 residues). Amino acid sequences were aligned with Muscle (MEGA5 software) and determination of the optimal amino acid substitution model and phylogenetic tree construction were done using MEGA5 software [45]. Neighbor-joining, maximum parsimony and maximum-likelihood phylogenetic trees of the individual

gene sequences were generated in MEGA5 by using the optimal model parameters and the option of complete deletion to eliminate positions containing gaps. Confidence levels for the branching points were determined using 1,000 bootstrap replicates.

Bioinformatics and statistical analysis

Searches for sequence similarity in the NCBI databases were carried out using BLAST algorithms [42]. Genome and nucleotide sequences were visualized and manipulated using the Artemis genome browser [46] and compared using ACT [47] in combination with WebACT [48]. The statistical analysis of incidence was performed by SAS9.2 software (SAS Institute Inc.) by Enterprise Guide 4.2 using generalized linear model analysis. The β -galactosidase and the necrotic area data were statistically analyzed using an analysis of variance, followed by Fisher's least significant difference test ($p = 0.05$), and for β -galactosidase activity on *P. protegens* Pf5, a Student's

t-test was carried out ($p = 0.05$), using the IBM.SSPS 19 software (IBM® Company).

Results

Involvement of *mbo* genes in mangotoxin production and virulence in *P. syringae* pv. *syringae* UMAF0158

Six mangotoxin deficient mutants of *P. syringae* pv. *syringae* UMAF0158, were previously obtained and characterized for mangotoxin production (Table 1 and Figure 1). Mangotoxin characterization showed that although these mutants did not show mangotoxin production, a slight production of a yet unknown antimicrobial compound was observed for mutants 4 β A2 (*mboB*) and 5 α C5 (*mboD*) (Figure 1). For two mutants (3 γ H1 and 6 γ F6), the Tn5 insertion was located in *mgoC* and *mgoA* respectively. Two other non-mangotoxin producing mutants were disrupted in the genes encoding the GacS/GacA two-component regulatory system (3 α E10 and 2 β B7 respectively). Growth of the *mgoA* mutant was shown to be similar to that of the

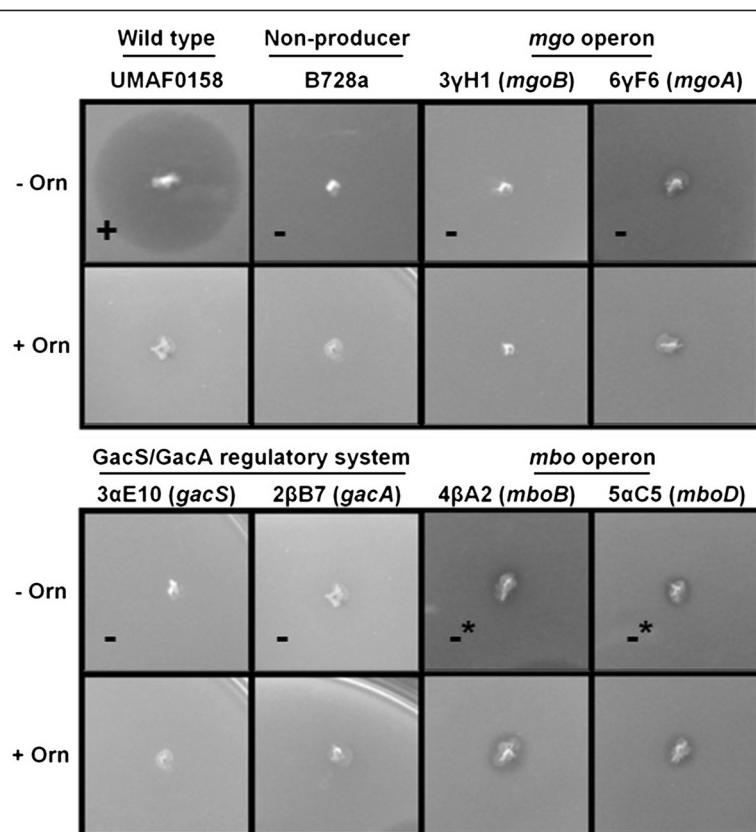
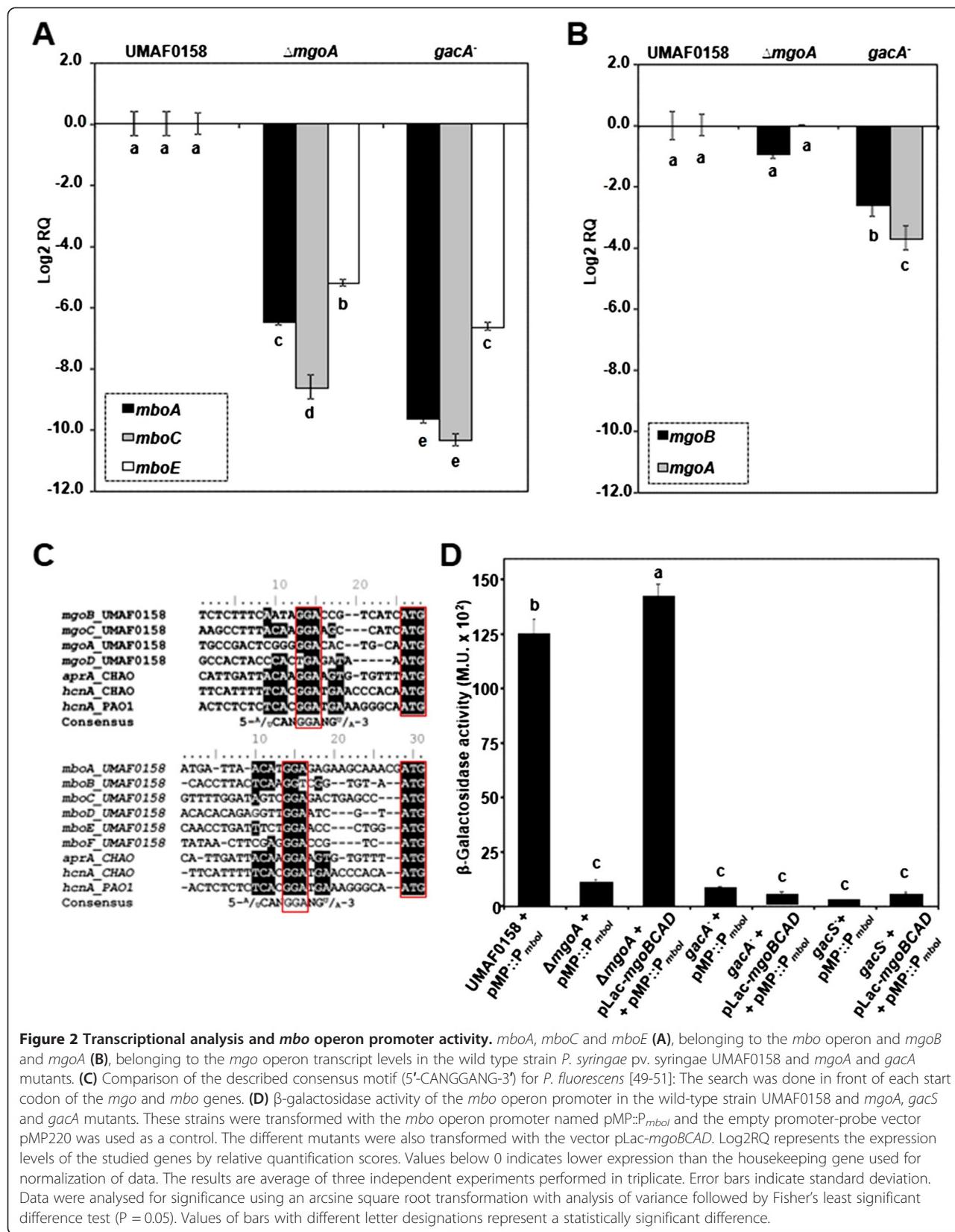


Figure 1 Mangotoxin production by random miniTn5 insertional mutants. Three pairs of mutants in different genes of the *mbo* and *mgo* operon, and in the *gacS/gacA* two-component regulatory system, obtained in previous works and tested for mangotoxin production. The corresponding disrupted gene is detailed in brackets. The *P. syringae* pv. *syringae* strains UMAF0158 (mangoxin-producing wild-type strain) and B728a (nonproducing) were used as references. Mangotoxin production was evaluated using PMS minimal medium supplemented or not with ornithine. The results are indicated as follows: - absence of inhibition halo, + presence of inhibition halo, -* slight toxicity which was not reverted by addition of ornithine. Toxic activity reverted in presence of ornithine denotes the production of mangotoxin.



wild type strain, with cell densities of up to 10^{11} cfu ml $^{-1}$ in liquid medium after 108 h of growth at 22°C (Additional file 2: Figure S1A). In contrast, the *gacA* mutant presented an altered growth, with cell densities in the stationary phase reaching only 10^9 cfu ml $^{-1}$ (Additional file 2: Figure S1A). The dynamics of the mangotoxin production in relation to bacterial growth was followed during four days of incubation. Mangotoxin production was detectable after 24 h of growth, increased up to 1.4 toxic units (T.U.), then reduced slightly upon entry of the stationary phase and then stabilized (Additional file 2: Figure S1B).

In order to know if the virulence of the derivative mutants *mboA*⁻ and *mgoA* was reduced in comparison with the wild type strain, detached tomato leaflets were artificially inoculated. Artificial inoculation experiments using detached tomato leaflets [4] showed that bacterial growth inside the tomato leaflets of the *mboA*⁻ and Δ *mgoA* mutants as well as their complemented derivatives followed similar dynamics (Additional file 3: Figure S2A). When inoculations were performed, development of necrotic lesions was observed on the leaf. Disease severity, represented by the necrotic area, showed that both mangotoxin defective mutants were less virulent than the wild type UMAF0158 (Additional file 3: Figure S2B and S2C). When derivative strains were complemented with the *mboA* and *mgoA* genes disease severity increased but complementation did not fully restore

virulence to wild type level (Additional file 3: Figure S2B and S2C).

Mangotoxin production and transcriptional regulation in the *gacA* and *mgoA* mutant

To study the role of *mgoA* and *gacA* in mangotoxin biosynthesis, transcription of the *mboACE* and *mgoBA* genes was analyzed for the wild type strain, and for the *mgoA* and *gacA* derivative mutants. Time course experiments showed that the *mbo* genes in the wild type are expressed at the highest level after 12 to 24 h (Additional file 4: Figure S3). Therefore all comparisons between wild type and mutants were performed at 18 h of growth. Transcript levels of the *mboACE* genes after 18 h of growth were significantly lower in the *gacA* and the *mgoA* mutants than in the wild type (Figure 2A). Also the transcript levels of *mgoB* and *mgoA* were significantly lower in the *gacA* mutant (Figure 2B). The *mgoA* mutation did not affect transcription of *gacS/gacA* (data not shown). Also *mboA*, *mboC*, or *mboE* mutations did not significantly affect transcription of *gacS/gacA* or *mgoA* (data not shown). These results indicate that the GacS/GacA two-component regulatory system affects transcription of both the *mbo* and *mgo* genes and that the product of the *mgo* operon influences transcription of the *mbo* genes. To further study if the GacS/GacA two-component regulatory system could regulate the *mgo* and *mbo* genes via RNA repressor binding proteins [49-51], the upstream

Table 2 Toxic activity of *P. syringae* pv *syringae* UMAF0158 mutants and *mgo* operon complemented strains

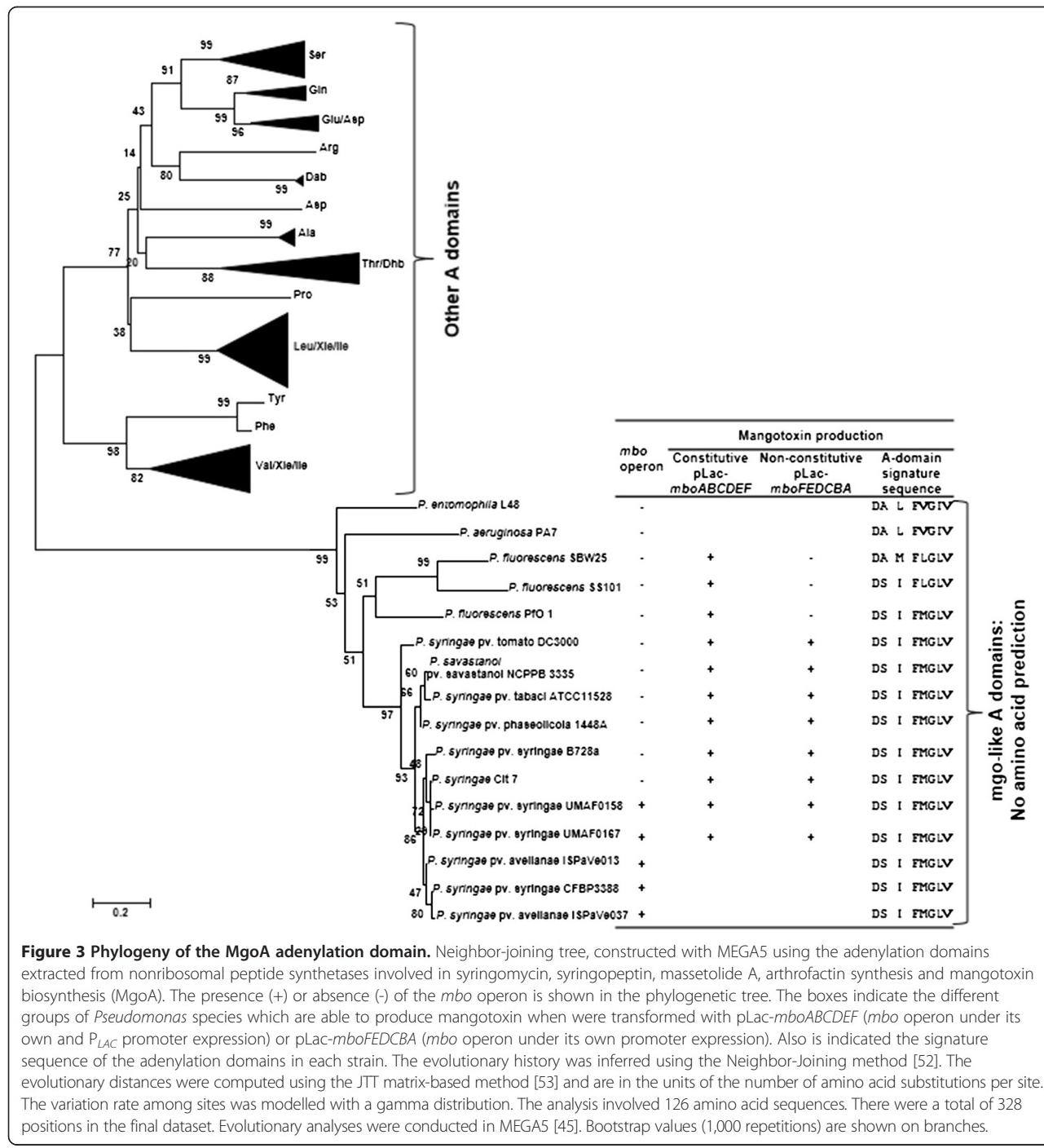
Strains	<i>E. coli</i> inhibition assay		Mangotoxin production
	PMS	PMS + ornithine	
Wild type strain and derivative mutants			
UMAF0158	+	-	Yes
<i>mboA</i> ⁻	-*	-*	No
Δ <i>mgoA</i>	-	-	No
<i>gacA</i> ⁻	-	-	No
<i>gacS</i> ⁻	-	-	No
Transformed with empty vector			
UMAF0158	+	-	Yes
<i>mboA</i> ⁻	-*	-*	No
Δ <i>mgoA</i>	-	-	No
<i>gacA</i> ⁻	-	-	No
<i>gacS</i> ⁻	-	-	No
Transformed with pLac-mgoBCAD			
UMAF0158	++	-	Yes
<i>mboA</i> ⁻	-*	-*	No
Δ <i>mgoA</i>	++	-	Yes
<i>gacA</i> ⁻	-	-	No
<i>gacS</i> ⁻	-	-	No

The results are indicated as follows: - absence of inhibition halo, + inhibition halo between 5-10 mm, ++ inhibition halo bigger 10 mm, -* slight toxicity which did not revert in presence of ornithine. Toxic activity, which reverts in the presence of ornithine, denotes the production of mangotoxin.

regions of the *mgo* and *mbo* genes were inspected for the presence of the described consensus motif (5'-CANG-GANG-3') previously described in *P. protegens* CHAO [49]. This motif allows the binding of the repressor to the RNA, and these repressor proteins can be removed by Gac/Rsm. The complete consensus sequence was not detected upstream of any of the *mbo/mgo* genes (Figure 2C). However, consensus GGA motifs for binding of the RNA binding proteins [49-51] were detected upstream of the *mbo*

and *mgo* operons (Figure 2C). It must be taken into account that the described consensus sequence is from *P. protegens* [49], and nothing is known yet about the recognition site of RNA binding proteins in *P. syringae*.

As the transcription of the *mgo* operon was substantially lower in the *gacA* mutant (Figure 2B), we subsequently tested whether introduction of extra copies of the *mgo* operon in the *gacS* or *gacA* mutant could restore mangotoxin production. When the *mgo* operon



was introduced in the *mgoA* mutant mangotoxin production was restored, which was not the case for the *mboA*, *gacA* and *gacS* mutants (Table 2).

The *mgo* operon is a positive regulator of *mbo* operon transcription

To further elucidate the role of the *mgo* operon in the regulation of mangotoxin biosynthesis, expression assays were carried out using a plasmid reporter construction consisting of the *mbo* operon promoter fused to a promoterless *lacZ* gene. When the plasmid reporter was transferred into the wild type strain, high levels of β-galactosidase activity were found, whereas for the *mgoA*, *gacA* and *gacS* mutants this activity was substantially lower (Figure 2D). For the *mgoA* mutant, complementation with the *mgo* operon restored β-galactosidase activity to similar levels as in the wild type strain (Figure 2D). In contrast, no restoration of the β-galactosidase activity was found when the *mgo* operon was introduced in the *gacS/gacA*, confirming results described above (Table 2).

MgoA phylogeny and mangotoxin production in other strains

The amino acid sequence of a typical non-ribosomal peptide synthetase (NRPS) displays an adenylation (A) domain responsible for recognition and subsequent activation of an amino acid substrate. It also contains the typical thiolation (T) and condensation (C) domains. Finally, the thioesterase (TE) domain releases the final molecule from the NRPS assembly line. Based on the specific signature sequences described previously for A domains, analysis of MgoA did not allow prediction of the amino acid to be activated. Therefore, a phylogenetic analysis was performed with multiple A domains from NRPSs of which activated amino acids are known and with MgoA from other *Pseudomonas* species (Figure 3 and Additional file 5: Figure S4). The results showed that the A domains from the different MgoA orthologues grouped in the same cluster, separate from other A domains for which the activated amino acid residue is known (Figure 3).

To determine if *mgoA* present in other *Pseudomonas* species can regulate the *mbo* operon, reporter constructs pLac-*mboABCDEF* (*mbo* operon under its own and under pLac promoter expression) and pLac-*mboFEDCBA* (*mbo* operon only under its own promoter expression) were used. Firstly, only specific *P. syringae* pathovars harbor the *mbo* operon, and almost all strains from these pathovars produce mangotoxin [29], with or without the introduction of the *mbo* operon containing plasmids (Figure 3). Our results showed that other *P. syringae* pathovars, that do not contain the *mbo* operon, are all able to produce mangotoxin when they were transformed with pLac-*mboABCDEF* and pLac-*mboFEDCBA* (Figure 3). When different *P. fluorescens* strains were transformed with

either vector, they only produced mangotoxin when the *mbo* operon was expressed constitutively but not when they were transformed with the *mbo* operon with its native promoter (Figure 3).

To further investigate if the *mgo* operon is able to regulate the expression of the *mbo* operon, we introduced the *mbo* operon promoter reporter construct (pMP::P_{mbo}) and the *mgo* genes in *P. protegens* Pf-5, which lacks both the *mgo* and the *mbo* operons in its genome. Compared to the promoter activity in the wild-type Pf-5 background, a two-fold increase in ectopic *mbo* promoter activity was observed when Pf-5 was complemented with the *mgo* operon (Figure 4A). When *P. protegens* Pf-5 was transformed with pLac-*mboABCDEF* (*mbo* operon under pLac regulation), it produces mangotoxin. However, when *P. protegens* Pf-5 was transformed with pMP-*mboFEDCBA* (*mbo* operon under only

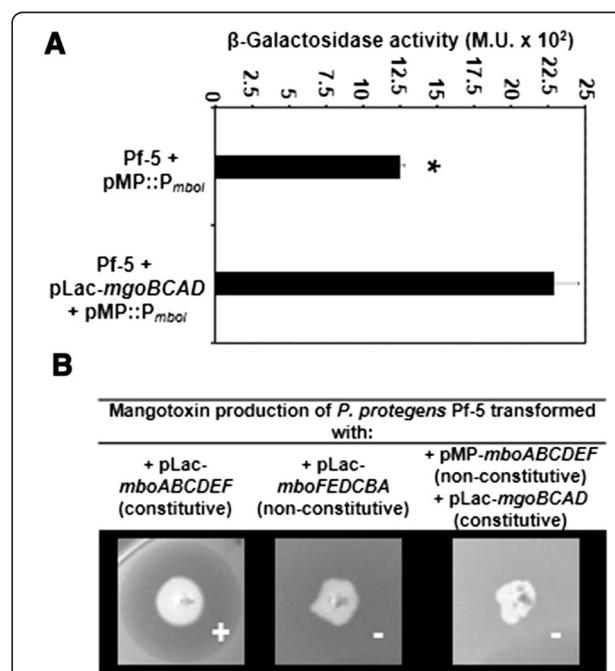


Figure 4 Heterologous expression and production of mangotoxin. **(A)** The *mbo* operon promoter activity in *P. protegens* Pf-5 transformed with the *mbo* operon promoter (pMP::P_{mbo}) and with the empty promoter-probe vector pMP220 was used as a control. To check the positive regulation of the *mgo* operon, the strain Pf-5 was transformed with the vector pLac-*mgoBCAD*. The result is the average of three independent experiments performed in triplicate. Error bars indicate standard deviation. **(B)** Mangotoxin production of *P. protegens* Pf-5 transformed with pLac-*mboABCDEF* (*mbo* operon under its own and P_{LAC} promoter expression), pLac-*mboFEDCBA* (*mbo* operon under its own promoter expression) and pLac-*mgoBCAD* (*mgo* operon under its own and P_{LAC} promoter expression) and pMP220-*mboABCDEF* (*mbo* operon under its own promoter expression). Data were analysed for significance using a Student's t-test (P = 0.05). Value of bar with an asterisk designation represent a statistically significant difference to the other bar value.

its own promoter expression) it was not able to produce detectable amounts of mangotoxin, neither in absence nor in presence of the *mgo* operon of *P. syringae* pv. *syringae* UMAF0158 (Figure 4B). Therefore, the presence of the *mbo* and *mgo* operons in *P. protegens* Pf-5 would be not sufficient for the production of detectable amounts of mangotoxin.

Discussion

The results of our study show that the regulation of mangotoxin biosynthesis in the plant pathogenic *P. syringae* pv. *syringae* strain UMAF0158 is governed by a complex interplay between the GacS/GacA two-component regulatory system, the nonribosomal peptide synthetase *mgoA* and the mangotoxin biosynthesis operon *mbo*. We showed that disruption of the *mbo* biosynthesis genes leads to reduced virulence. Introduction of the *mbo* operon in these biosynthesis mutants restored mangotoxin production but did not lead to full restoration of virulence on tomato leaflets. Multiple copies of the plasmid with the *mbo* operon could lead to overproduction of mangotoxin which may affect the regulation or production of other virulence factors such as syringomycin and syringopeptin.

Taken together the obtained results of this work and the previously described data [4,6,7], a simplified model for the interplay among these genes can be constructed (Figure 5). In this model, the GacS/GacA two-component regulatory

system receives a yet unknown signal that activates a set of small RNAs [8,50,54]. The expression of genes regulated by the GacS/GacA might be mediated through the Rsm pathway [55,56]. In fact, components of this pathway such as the three small RNAs RsmX, RsmY and RsmZ and two RNA-binding proteins (RsmA and RsmE) were found in the genome of *P. syringae* pv. *syringae* UMAF0158 (Unpublished data). Transcriptional analysis of the *mgo*, *mbo* and *gac* genes showed that the *mbo* genes were markedly down-regulated in both the *gacA* and *mgoA* mutants. On the other hand, the transcriptional levels of *mgoB* and *mgoA*, also showed down-regulation in the *gacA* mutant, indicating that the *mgo* operon is also under regulation by the GacS/GacA two-component regulatory system. These data suggest that GacS/GacA is regulating the *mbo* operon expression via the *mgo* operon, however direct regulation of the *mbo* operon by the two-component regulatory system *gacS/gacA* cannot be excluded (Figure 5).

Transcriptional analysis with a *lacZ* fusion on the promoter of the *mbo* operon (P_{mbo}), revealed that the product of the *mgo* operon could act as positive regulator of *mbo* transcription. Interestingly, the *pvfC* gene (homologue of *mgoA*) is considered a regulator of virulence in *P. entomophila*, but appears not to be part of the GacS/GacA regulatory cascade [28]. In strain UMAF0158, introduction of the *mgo* operon in a *gac* mutant could not restore mangotoxin production or *mbo*-promoter activity, suggesting that next

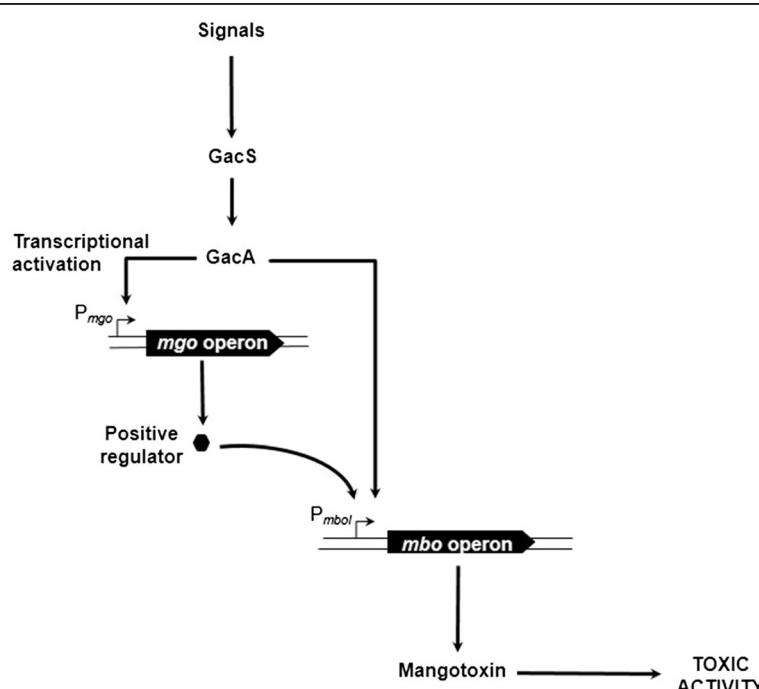


Figure 5 Proposed model for regulation of mangotoxin biosynthesis in *P. syringae* pv. *syringae*. In this model, GacS/GacA two-component regulatory system activates directly or indirectly the transcription of the *mgo* operon. And the *mgo* operon could synthesize a positive regulator of the *mbo* operon transcription. The *mbo* operon produces mangotoxin which acts as virulence factor.

to the *mgo* operon, additional factors are regulated by the *gac* system that influence mangotoxin production. It is worth noting that *P. entomophila* and *P. syringae* pv. *syringae* harbor two different genetic backgrounds, adapted to different environments. The first is found in diverse environments such as soil, aquatic ecosystems, rhizosphere, and in pathogenic interactions with *Drosophila melanogaster* [57]. The second is adapted for plant infection and epiphytic survival [3]. Therefore, the regulatory roles of these orthologues can substantially differ between these two *Pseudomonas* species. On the other hand, the fact that both PvfC and MgoA are involved in the regulation of virulence could indicate that in other *Pseudomonas* spp. these factors would be involved in the regulation of virulence and/or secondary metabolite production.

Phylogenetic analysis of MgoA and the adenylation domains suggested an evolutionary specialization of this protein into the *Pseudomonas* genus. In this context, it is worth noting that the transformation of the *mbo* operon under the expression of its own promoter only confers mangotoxin production in the *P. syringae* group and not in the *P. fluorescens* group. Therefore, it seems that the NRPS MgoA is involved in different signal transduction pathways depending of the *Pseudomonas* species. In the case of *P. syringae*, MgoA appears to activate mangotoxin production. It remains to be studied if MgoA is also involved in the regulation and production of other antimetabolites in the *P. syringae* group, such as tabtoxin and phaseolotoxin. The positive regulation of the *mbo* operon promoter activity in the presence of the *mgo* operon in Pf-5, combined with the lack of detectable amounts of mangotoxin suggests that additional factors for mangotoxin biosynthesis or its export are not present in the *P. fluorescens* group.

Conclusions

In summary, for *P. syringae* pv. *syringae* UMAF0158, the GacS/GacA two-component system regulates transcription of the *mgo* and *mbo* operons and thereby mangotoxin biosynthesis. At the same time, the *mgo* operon product seems to act as a positive regulator of the *mbo* operon. The proposed model for mangotoxin biosynthesis is a simplified and initial overview of the interaction between the *gac*, *mgo* and *mbo* gene products based on the results obtained in the current study. This is the first evidence of the interplay between MgoA and the GacS/GacA two-component regulatory system in the regulation of the mangotoxin biosynthesis.

Ethics statement

We the authors hereby declare that the research performed with plants has been conducted in accordance with institutional, national and international guidelines.

Additional files

Additional file 1: Table S1. Primers used in this study.

Additional file 2: Figure S1. Growth characteristics of *P. syringae* pv. *syringae* strain UMAF0158 and the derivatives *mgoA* and *gacA* mutants. (A) Growth of the wild type strain UMAF0158 and the *mgoA* ($\Delta mgoA$) and *gacA* (2 β B7) mutants at 22°C in PMS. At each time point, the bacterial density was estimated by serial dilutions and colony counts on plates of selective medium and expressed as log cfu ml⁻¹ of culture. (B) UMAF0158 mangotoxin production at 22°C in PMS. At each time point, the mangotoxin production was estimated using cell-free filtrate and represented as the previously defined toxic units (T.U.). The dashed line represents the detection limit of the technique. Mean values for three replicates are given; the error bars represent the standard errors of the mean.

Additional file 3: Figure S2. Virulence analysis of the wild type strain *P. syringae* pv. *syringae* UMAF0158 and corresponding derivatives using a detached tomato leaf assay. (A) *In planta* growth inside the tomato leaflets after H₂O₂ surface disinfection of the wild type strain UMAF0158, *mgoA* and *mboA* mutants, and their respective complemented derivatives. (B) Severity of necrotic symptoms (necrotic area) on tomato leaflets inoculated with wild type strain UMAF0158, the mutants in *mboA* and *mgoA* with their respective complemented derivatives. The total necrotic area (mm²) from 30 inoculated points on tomato leaflets was measured 10 days after inoculation and used to compare the severity of necrotic symptoms produced by the different strains. (C) Representative pictures of the necrotic lesions produced by the wild type strain and the different mutants at 10 dpi. Different letters denote statistically significant differences at *p* = 0.05, according to analysis of variance followed by Fisher's least significant difference test.

Additional file 4: Figure S3. *mboACE* transcript levels in the wild type strain UMAF0158. Relative expression of the genes involved in the mangotoxin biosynthesis at the different time points during the growth curve. For each time point, mean values of four biological replicates are given; the error bars represent the standard errors of the mean.

Additional file 5: Figure S4. Phylogenetic analysis of the MgoA of different *Pseudomonas* spp. Neighbor-joining tree was constructed with MEGA5 using a partial sequence of MgoA. The boxes indicate the different groups of *Pseudomonas* and the presence (*mbo* +) or absence (*mbo* -) of the *mbo* operon. The evolutionary history was inferred using the Neighbor-Joining method [52]. The evolutionary distances were computed using the JTT matrix-based method [53] and are in the units of the number of amino acid substitutions per site. The rate variation among sites was modelled with a gamma distribution. The analysis involved 126 amino acid sequences. There were a total of 1015 positions in the final dataset. Evolutionary analyses were conducted in MEGAS [45]. *Burkholderia cepacia* J2315 was used as the outgroup. Bootstrap values (1,000 repetitions) are shown on the branches.

Competing interests

We the authors hereby declare that there is no conflict of interests concerning this manuscript.

Authors' contributions

VJC, MV, EA, AV, JMR and FMC conceived the study. VJC and EA did all the cloning and genetics of this study. VJC and MV did the Q-PCR experiments and analysis. VJC and JAG did complementation and reporter construct experiments. JMR and AV supported the research. VJC, MV, JMR and FMC wrote the manuscript. VJC, EA, MV, AV, JMR and FMC coordinated and critically revised the manuscript. All authors read and approved the manuscript.

Acknowledgements

This work was supported by grants from the Regional Government of Andalucía (Spain), grants from CICE - Junta de Andalucía, Ayudas Grupo PAIDI AGR-169, and Proyecto de Excelencia (P07-AGR-02471) and Plan Nacional de I + D + I del Ministerio de Ciencia e Innovación (AGL2011-30354-C02-01) cofinanced by FEDER (EU). Plan Propio of the University of Málaga funded a stay of VJC in the Wageningen University, The Netherlands. This is publication No. 5584 of the Netherlands Institute of Ecology (NIOO-KNAW). VJC was

supported with a fellowship from Junta de Andalucía, Spain, and EA with a JAEDoc grant from the CSIC, which was co-financed by ESF. We also thank Cayo Ramos and his group for their help in this research.

Author details

¹Departamento de Microbiología, Facultad de Ciencias, Instituto de Hortofruticultura Subtropical y Mediterránea "La Mayora"-Universidad de Málaga-Consejo Superior de Investigaciones Científicas (IHSM-UMA-CSIC), Universidad de Málaga, Campus de Teatinos, 29071 Málaga, Spain. ²Instituto de Hortofruticultura Subtropical y Mediterránea "La Mayora"-Universidad de Málaga-Consejo Superior de Investigaciones Científicas (IHSM-UMA-CSIC), Estación Experimental La Mayora, Algarrobo-Costa, 29750 Málaga, Spain.

³Laboratory of Phytopathology, Wageningen University, 6708 PB Wageningen, The Netherlands. ⁴Department of Microbial Ecology, The Netherlands Institute of Ecology (NIOO-KNAW), Wageningen, The Netherlands. ⁵BIMERIT Research Centre, School of Microbiology, University College Cork, National University of Ireland, Cork, Ireland.

Received: 10 December 2013 Accepted: 12 February 2014

Published: 21 February 2014

References

1. Kennelly MM, Cazorla FM, de Vicente A, Ramos C, Sundin GW: *Pseudomonas syringae* diseases of fruit trees: progress toward understanding and control. *Plant Dis* 2007, **91**(1):4–17.
2. Arrebola E, Cazorla FM, Durán VE, Rivera E, Olea F, Codina JC, Pérez-García A, de Vicente A: Mangotoxin: a novel antimetabolite toxin produced by *Pseudomonas syringae* inhibiting ornithine/arginine biosynthesis. *Physiol Mol Plant Path* 2003, **63**(3):117–127.
3. Cazorla FM, Torés JA, Olalla L, Pérez-García A, Farré JM, de Vicente A: Bacterial apical necrosis of mango in southern Spain: a disease caused by *Pseudomonas syringae* pv. *syringae*. *Phytopathology* 1998, **88**(7):614–620.
4. Arrebola E, Cazorla FM, Romero D, Pérez-García A, de Vicente A: A nonribosomal peptide synthetase gene (*mgoA*) of *Pseudomonas syringae* pv. *syringae* is involved in mangotoxin biosynthesis and is required for full virulence. *Mol Plant-Microbe Interact* 2007, **20**(5):500–509.
5. Arrebola E, Cazorla FM, Codina JC, Gutiérrez-Barranquero JA, Pérez-García A, de Vicente A: Contribution of mangotoxin to the virulence and epiphytic fitness of *Pseudomonas syringae* pv. *syringae*. *Int Microbiol* 2009, **12**(1139–6709):87–95.
6. Carrión VJ, Arrebola E, Cazorla FM, Murillo J, de Vicente A: The *mbo* operon is specific and essential for biosynthesis of mangotoxin in *Pseudomonas syringae*. *PLoS One* 2012, **7**(5):e36709.
7. Arrebola E, Carrión VJ, Cazorla FM, Pérez-García A, Murillo J, de Vicente A: Characterisation of the *mgo* operon in *Pseudomonas syringae* pv. *syringae* UMAF0158 that is required for mangotoxin production. *BMC Microbiol* 2012, **12**(1):10.
8. Heeb S, Haas D: Regulatory roles of the GacS/GacA two-component system in plant-associated and other gram-negative bacteria. *Mol Plant-Microbe Interact* 2001, **14**(12):1351–1363.
9. Chancery ST, Wood DW, Pierson LS: Two-component transcriptional regulation of N-acyl-homoserine lactone production in *Pseudomonas aureofaciens*. *Appl Environ Microbiol* 1999, **65**(6):2294–2299.
10. Kay E, Humair B, Déneraud V, Riedel K, Spahr S, Eberl L, Valverde C, Haas D: Two GacA-dependent small RNAs modulate the quorum-sensing response in *Pseudomonas aeruginosa*. *J Bacteriol* 2006, **188**(16):6026–6033.
11. Corbell N, Loper JE: A global regulator of secondary metabolite production in *Pseudomonas fluorescens* Pf-5. *J Bacteriol* 1995, **177**(21):6230–6236.
12. Whistler CA, Pierson LS III: Repression of phenazine antibiotic production in *Pseudomonas aureofaciens* strain 30-84 by RpeA. *J Bacteriol* 2003, **185**(13):3718–3725.
13. Hassan KA, Johnson A, Shaffer BT, Ren Q, Kidarsa TA, Elbourne LDH, Hartney S, Duboy R, Goebel NC, Zabriskie TM, Paulsen IT, Loper JE: Inactivation of the Gac response regulator in *Pseudomonas fluorescens* Pf-5 has far-reaching transcriptomic consequences. *Environ Microbiol* 2010, **12**(4):899–915.
14. Cheng X, de Bruijn I, van der Voort M, Loper JE, Raaijmakers JM: The Gac regulon of *Pseudomonas fluorescens* SBW25. *Environ Microbiol Rep* 2013, **5**(4):608–619.
15. Parkins MD, Cieri H, Storey DG: *Pseudomonas aeruginosa* GacA, a factor in multihost virulence, is also essential for biofilm formation. *Mol Microbiol* 2001, **40**(5):1215–1226.
16. Petrova OE, Sauer K: The novel two-component regulatory system BfISR regulates biofilm development by controlling the small RNA *rsmZ* through CafA. *J Bacteriol* 2010, **192**(20):5275–5288.
17. Muller J, Shukla S, Jost K, Spormann A: The *mxd* operon in *Shewanella oneidensis* MR-1 is induced in response to starvation and regulated by ArcS/ArcA and BarA/UvrY. *BMC Microbiol* 2013, **13**(1):119.
18. Lu S-E, Scholz-Schroeder BK, Gross DC: Characterization of the *salA*, *syrF*, and *syrG* regulatory genes located at the right border of the syringomycin gene cluster of *Pseudomonas syringae* pv. *syringae*. *Mol Plant-Microbe Interact* 2002, **15**(1):43–53.
19. Wang N, Lu S-E, Wang J, Chen ZJ, Gross DC: The expression of genes encoding lipopeptide phytotoxins by *Pseudomonas syringae* pv. *syringae* is coordinated in response to plant signal molecules. *Mol Plant-Microbe Interact* 2006, **19**(3):257–269.
20. Lu S-E, Wang N, Wang J, Chen ZJ, Gross DC: Oligonucleotide microarray analysis of the *SalA* regulon controlling phytotoxin production by *Pseudomonas syringae* pv. *syringae*. *Mol Plant-Microbe Interact* 2005, **18**(4):324–333.
21. Barta TM, Kinscherf TG, Willis DK: Regulation of tabtoxin production by the *lemA* gene in *Pseudomonas syringae*. *J Bacteriol* 1992, **174**(9):3021–3029.
22. Bender CL, Alarcón-Chaidez F, Gross DC: *Pseudomonas syringae* phytotoxins: mode of action, regulation, and biosynthesis by peptide and polyketide synthetases. *Microbiol Mol Biol Rev* 1999, **63**(2):266–292.
23. de la Torre-Zavala S, Aguilera S, Ibarra-Laclette E, Hernandez-Flores JL, Hernández-Morales A, Murillo J, Alvarez-Morales A: Gene expression of Pht cluster genes and a putative non-ribosomal peptide synthetase required for phaseolotoxin production is regulated by GacS/GacA in *Pseudomonas syringae* pv. *phaseolicola*. *Res Microbiol* 2011, **162**(5):488–498.
24. Willis DK, Hrabak EM, Rich JJ, Barta TM, Lindow SE, Panopoulos NJ: Isolation and characterization of a *Pseudomonas syringae* pv. *syringae* mutant deficient in lesion formation on bean. *Mol Plant-Microbe Interact* 1990, **3**(3):149–156.
25. Chatterjee A, Cui Y, Yang H, Collmer A, Alfano JR, Chatterjee AK: GacA, the response regulator of a two-component system, acts as a master regulator in *Pseudomonas syringae* pv. *tomato* DC3000 by controlling regulatory RNA, transcriptional activators, and alternate sigma factors. *Mol Plant-Microbe Interact* 2003, **16**(12):1106–1117.
26. Lindeberg M, Myers CR, Collmer A, Schneider DJ: Roadmap to new virulence determinants in *Pseudomonas syringae*: insights from comparative genomics and genome organization. *Mol Plant-Microbe Interact* 2008, **21**(6):685–700.
27. Loper JE, Hassan KA, Mavrodin DV, Davis EW II, Lim CK, Shaffer BT, Elbourne LDH, Stockwell VO, Hartney SL, Breakwell K, Henkels MD, Tetu SG, Rangel LJ, Kidarsa TA, Wilson NL, van de Mortel JE, Song C, Blumhagen R, Radune D, Hostetler JB, Brinkac LM, Durkin AS, Kluepfel DA, Wechter WP, Anderson AJ, Kim YC, Pierson LS III, Pierson EA, Lindow SE, Kobayashi DY, et al: Comparative genomics of Plant-Associated *Pseudomonas* spp.: insights into diversity and inheritance of traits involved in multitrophic interactions. *PLoS Genet* 2012, **8**(7):e1002784.
28. Vallet-Gely I, Opota O, Boniface A, Novikov A, Lemaitre B: A secondary metabolite acting as a signalling molecule controls *Pseudomonas entomophila* virulence. *Cell Microbiol* 2010, **12**(11):1666–1679.
29. Carrión VJ, Gutiérrez-Barranquero JA, Arrebola E, Bardaji L, Codina JC, de Vicente A, Cazorla FM, Murillo J: The mangotoxin biosynthetic operon (*mbo*) is specifically distributed within *Pseudomonas syringae*. *Genospecies 1* was acquired only once during evolution. *Appl Environ Microbiol* 2013, **79**(3):756–767.
30. Gutiérrez-Barranquero JA, Carrión VJ, Murillo J, Arrebola E, Arnold DL, Cazorla FM, de Vicente A: A *Pseudomonas syringae* diversity survey reveals a differentiated phylotype of the pathovar *syringae* associated with the mango host and mangotoxin production. *Phytopathology* 2013, **103**(11):1115–1129.
31. King EO, Ward MK, Raney DE: Two simple media for the demonstration of pyocyanin and fluorescin. *J Lab Clin Med* 1954, **44**(2):301–307.
32. Gasson MJ: Indicator technique for antimetabolic toxin production by phytopathogenic species of *Pseudomonas*. *Appl Environ Microbiol* 1980, **39**(1):25–29.
33. Hanahan D: Studies on transformation of *Escherichia coli* with plasmids. *J M Biol* 1983, **166**(4):557–580.
34. Feil H, Feil WS, Chain P, Larimer F, DiBartolo G, Copeland A, Lykidis A, Trong S, Nolan M, Gotsman E, Thiel J, Malfatti S, Loper JE, Lapidus A, Detter JC, Land M, Richardson PM, Kyrpides NC, Ivanova N, Lindow SE: Comparison of the complete genome sequences of *Pseudomonas syringae* pv. *syringae* B728a and pv. *tomato* DC3000. *Proc Natl Acad Sci USA* 2005, **102**(31):11064–11069.

35. Howell CR, Stipanovic RD: Suppression of *Pythium ultimum*-induced damping-off of cotton seedlings by *Pseudomonas fluorescens* and its antibiotic, pyoluteorin. *Phytopathology* 1980, 70:712–715.
36. Kovach ME, Elzer PH, Steven Hill D, Robertson GT, Farris MA, Roop li RM, Peterson KM: Four new derivatives of the broad-host-range cloning vector pBBR1MCS, carrying different antibiotic-resistance cassettes. *Gene* 1995, 166(1):175–176.
37. Spaink HP, Okker RJH, Wijffelman CA, Pees E, Lugtenberg BJJ: Promoters in the nodulation region of the *Rhizobium leguminosarum* Sym plasmid pRL1JI. *Plant Mol Biol* 1987, 9(1):27–39.
38. Sambrook J, Russel DW: *Molecular Cloning: A Laboratory Manual*. Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press; 2001.
39. de Bruijn I, de Kock MJD, de Waard P, van Beek TA, Raaijmakers JM: Masetolide A biosynthesis in *Pseudomonas fluorescens*. *J Bacteriol* 2008, 190(8):2777–2789.
40. de Bruijn I, Raaijmakers JM: Regulation of cyclic lipopeptide biosynthesis in *Pseudomonas fluorescens* by the ClpP protease. *J Bacteriol* 2009, 191(6):1910–1923.
41. Miller JH: *Experiments in Molecular Genetics*. Cold Spring Harbor, NY: Cold Spring Harbor Laboratory; 1972.
42. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ: Basic local alignment search tool. *J Mol Biol* 1990, 215(3):403–410.
43. Bachmann BO, Ravel J: Chapter 8 methods for *in silico* prediction of microbial polyketide and nonribosomal peptide biosynthetic pathways from DNA sequence data. In *Method Enzymol*. Edited by David AH. PA: Academic Press; 2009:181–217. vol. Volume 458.
44. de Bruijn I, de Kock MJD, Yang M, de Waard P, van Beek TA, Raaijmakers JM: Genome-based discovery, structure prediction and functional analysis of cyclic lipopeptide antibiotics in *Pseudomonas* species. *Mol Microbiol* 2007, 63(2):417–428.
45. Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S: MEGA5: Molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Mol Biol Evol* 2011, 28(10):2731–2739.
46. Rutherford K, Parkhill J, Crook J, Horsnell T, Rice P, Rajandream MA, Barrell B: Artemis: sequence visualization and annotation. *Bioinformatics* 2000, 16(10):944–945.
47. Carver TJ, Rutherford KM, Berriman M, Rajandream MA, Barrell BG, Parkhill J: ACT: the Artemis comparison tool. *Bioinformatics* 2005, 21(16):3422–3423.
48. Abbott JC, Aanensen DM, Rutherford K, Butcher S, Spratt BG: WebACT—an online companion for the Artemis Comparison Tool. *Bioinformatics* 2005, 21(18):3665–3666.
49. Blumer C, Heeb S, Pessi G, Haas D: Global GacA-steered control of cyanide and exoprotease production in *Pseudomonas fluorescens* involves specific ribosome binding sites. *Proc Natl Acad Sci USA* 1999, 96(24):14073–14078.
50. Valverde C, Lindell M, Wagner EGH, Haas D: A repeated GGA motif is critical for the activity and stability of the riboregulator RsmY of *Pseudomonas fluorescens*. *J Biol Chem* 2004, 279(24):25066–25074.
51. Dubey AK, Baker CS, Suzuki K, Jones AD, Pandit P, Romeo T, Babitzke P: CsrA regulates translation of the *Escherichia coli* carbon starvation gene, cstA, by blocking ribosome access to the cstA transcript. *J Bacteriol* 2003, 185(15):4450–4460.
52. Saitou N, Nei M: The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Mol Biol Evol* 1987, 4(4):406–425.
53. Jones DT, Taylor WR, Thornton JM: The rapid generation of mutation data matrices from protein sequences. *Comput Appl Biosci* 1992, 8(3):275–282.
54. Lapouge K, Sineva E, Lindell M, Starke K, Baker CS, Babitzke P, Haas D: Mechanism of hcnA mRNA recognition in the Gac/Rsm signal transduction pathway of *Pseudomonas fluorescens*. *Mol Microbiol* 2007, 66(2):341–356.
55. Lapouge K, Schubert M, Allain FHT, Haas D: Gac/Rsm signal transduction pathway of gamma-proteobacteria: from RNA recognition to regulation of social behaviour. *Mol Microbiol* 2008, 67(2):241–253.
56. Kay E, Dubuis C, Haas D: Three small RNAs jointly ensure secondary metabolism and biocontrol in *Pseudomonas fluorescens* CHA0. *Proc Natl Acad Sci USA* 2005, 102(47):17136–17141.
57. Vodovar N, Vallenet D, Cruveiller S, Rouy Z, Barbe V, Acosta C, Cattolico L, Jubin C, Lajus A, Segurens B, Vacherie B, Wincker P, Weissenbach J, Lemaitre B, Médigue C, Boccard F: Complete genome sequence of the entomopathogenic and metabolically versatile soil bacterium *Pseudomonas entomophila*. *Nat Biotechnol* 2006, 24(6):673–679.

doi:10.1186/1471-2180-14-46

Cite this article as: Carrión et al.: Mangotoxin production of *Pseudomonas syringae* pv. *syringae* is regulated by MgoA. *BMC Microbiology* 2014 14:46.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

